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Therapeutic Efficacy of Phytopreparation for the Prevention and Treatment of Varroosis and Nosemosis of Honey Bees in Western Kazakhstan

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ABSTRACT

Bee diseases significantly hinder the development of the beekeeping industry and reduce its profitability. The use of various chemical origin preparations for treating bee diseases ensures that the infestation of bee colonies is kept to a minimum. However, the preparations have a negative impact on the bee body and contaminate bee products. Therefore, this study aimed to investigate the therapeutic efficacy of a new phytopreparation in the conditions of West Kazakhstan region. The phytopreparation was developed using medicinal plants native to the Republic of Kazakhstan. The therapeutic dose of phytopreparation (10 and 15mL/1L of 20% and 50% sugar syrup) was established using the laboratory method of dosed feeding. The research and production experiment demonstrated that the phytopreparation exhibits a pronounced anti-varroosis and anti-nosemosis effect. In the treatment of varroosis, the intensity of invasion is reduced to a safe level when used by the method of feeding at a dose of 10-15mL/1L sugar syrup in the spring period, the therapeutic efficiency is 86-85.7%, when irrigated in the specified doses - 84.3-85%. In the treatment of nosemosis, the preparation's effectiveness reaches 100%. The preparation can also be used for complex treatments and systematic therapeutic and preventive protection of bee colonies, as well as to stimulate the development of healthy bee colonies, an increasing their productivity.

Keywords: Honey bee, Varroosis, Nosemosis, Treatment, Phytopreparation

INTRODUCTION

The honey bee (*Apis mellifera*) plays a key role in biodiversity conservation and provides several unique products for the food industry and pharmaceuticals. Therefore, beekeeping is an important component of every country's economy.

Despite the annual increase in the number of bee colonies, beekeeping in many countries faces challenges that hinder the industry's growth (Wakgari et al., 2021). In Kazakhstan, as well as worldwide, one such factor is bee diseases, among which the most widespread and dangerous are varroa mites and nosemosis. Currently, these diseases are widespread, affecting between 10 and 100% of bee colonies (Chemurot et al., 2016; Tokarev et al., 2018; Baigazanov et al., 2022; Robi et al., 2023; Bava et al., 2023; Cilia et al., 2023; Farida et al., 2024). They adversely affect

the bee organism, causing a decrease in productivity and the death of bee colonies, which results in significant damage to beekeeping.

The Varroa mite is the most significant pest for bee colonies, as it affects them at all stages of development (Noël et al., 2020). Against the background of Varroa infestation, there is a tendency to the growth of infections (ascospherosis, American and European rot), the causative agents of which are transmitted by Varroa mite (Mikhaltsevich & Velichko, 1996; Grobov et al., 2008; Reams & Rangel, 2022; Dequenne et al., 2022).

The negative impact of *Nosema spp.* on bees is also significant. It is known that already 14 days after the development of *N. ceranae* in the middle intestine of bees, the amount of lipids in the fat body of honey bees decreases sharply. Especially strongly (up to 50%) lipid reserves decrease in the fall period (Gilbert et al., 2024).

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Additionally, bees infected with *N. ceranae* are protein deficient due to midgut lesions, resulting in a lower survival rate of the bees (Panek et al., 2018), which can lead to weakening and death of the bee colony (Mayack & Naug, 2009; Bekele, 2015; Ostap-Chec et al., 2024).

Various methods are used to protect bee colonies from disease, but the most effective is the use of medicines. It should be noted that most of the preparations used in beekeeping are products of biological and chemical production (Formato et al., 2011; Vandervalk et al., 2014; Bava et al., 2023; Aurell et al., 2024; Jack et al., 2024). They are often expensive or unavailable to beekeepers (especially in remote areas) and their choice is limited, as they are imported to the Republic of Kazakhstan from other countries. In some countries, the use of these preparations is prohibited or restricted (Chaimanee et al., 2021; Formato et al., 2022).

The uncontrolled and illiterate use of improvised veterinary and medical preparations causes even more damage, as it results in the death of bee colonies, the emergence of resistant species of pathogens, and outbreaks of bee diseases not previously encountered in a particular region. Additionally, it impacts the ecology of bee products that contain by-product compounds (Formato et al., 2011; Guün et al., 2024).

Studies have confirmed that chemical origin preparations negatively affect bees by accumulating in their bodies (Shakarian, 1981; Sotnikov, 1982; Terpin et al., 2019; Ahmad & Elsaiegh, 2022), also affect the exterior traits of honeybees, (Larkina & Lapynina, 2021), accumulate in honeycombs, honey (Er Demirhan & Demirhan, 2022), perga (Lambert et al., 2013; Lozano et al., 2019) and persist in them for a long time (more than 6 months) (Formato et al., 2011). Honey contaminated with veterinary preparation residues may acquire undesirable properties, including allergenicity, carcinogenicity, effects on the reproductive system, and teratogenicity (Rodrigues et al., 2024; Shoaei et al., 2024).

Scientists in many countries worldwide are conducting research on the development of new biologically active, therapeutic, and prophylactic preparations that are safe for both bees and humans (Formato et al., 2011; Kunat-Budzynńska et al., 2022; El-Seedi et al., 2022). Compounds found in plants are gaining attention because they exhibit antimicrobial, antifungal, antiviral and acaricidal activities (Bava et al., 2023). When isolated from plants, these compounds are able to degrade over time when exposed to light, air, and moisture. In addition, plant-based preparations have a balanced chemical composition and have combined and targeted therapeutic and biostimulatory actions. There is evidence from various sources on the use of more than 250 species of medicinal plants for the prevention and treatment of bee diseases (Kerimaliev, 2005; Khan et al., 2019; Pasca et al., 2021; El-Sayed et al., 2024), to stimulate the development and increase the productivity of bee colonies.

Medicinal plants are used in various dosage forms, including extracts, fumigants, and essential oils. Of these, extracts have the greatest therapeutic effect. Different extracts of the same plant inhibit pathogen development, but ethanolic extracts have the best results in most cases (Formato et al., 2022). Plant essential oils are used as impregnated strips on veneer and cardboard, or by spraying them on frames with bees (Rashidet et al., 2020). The efficacy of such treatment under field conditions for nosema can range from 71.21 to 84% (Ozuicli et al., 2023; Bava et al., 2023; Boonmee et al., 2024). For varroosis it can be as high as 97% (Mahmood et al., 2014; Salkova et al., 2024).

Fungi also have the potential to mitigate the negative effects of environmental factors. For use, of which only one contains medicinal plants, three are a combination of a chemical drug with essential oils of plants, and 8 are of chemical origin. Therefore, the purpose of this research is to investigate the therapeutic efficacy of a new phytopreparation for the prevention and treatment of varroosis and nosomosis in honeybees in the Western Kazakhstan region.

MATERIALS & METHODS

Ethical Approval

The experiment plan and handling of bees in the current study were approved by the local Commission on Biological Ethics of «Batys Zoo Vet Service» LLP (Protocol No. 276, dated 30.04.2024). When conducting experiments, we were guided by the recommendations of the Board of the Eurasian Economic Commission (2023), as outlined in the guidelines for working with laboratory (experimental) animals when conducting preclinical (non-clinical) studies.

Place of Study

The study was conducted by the Zhangir Khan West -Kazakhstan Agrarian – Technical University based on the beekeeping farm «Golden Beehive» located in the Michurinsk rural district of Bayterek (Fig. 1). The area of the land plot for the apiary is 5.5 hectares. The farm has about 100 bee colonies of the Carpathian breed, which are kept in wooden Dadan hives. The apiary is mobile; in winter, the hives are kept in wintering houses; in the spring-summerautumn season, they are placed near the sources of honey collection (crop fields, meadows, forests).

Phytopreparation

To obtain a phytopreparation for the prevention and complex treatment of bee varroosis and nosemosis, the following medicinal plants were taken: yarrow (Achillea millefolium), bitter wormwood (Artemisia absinthium), common pine (buds) (Pinus sylvestris). The finished preparation is a 10% extract of plant material in 70% ethanol (for 1 L of ethanol, take 50g of yarrow, 40g of wormwood, and 10g of pine buds). It is a brown-green liquid with a bitter taste and herbal aroma. All plants used to produce the preparation were collected in the region and identified by specialist N.V. Valitova. All plants for the preparation were collected on the territory of the region by the rules for the use of plant life as approved by order of the Council of the Eurasian Economic Commission (2018) and Acting Minister of Ecology and Natural Resources of the Republic of Kazakhstan (2023) on approval of the Rules of proper practice of cultivation, collection, processing, and storage of initial raw materials of plant origin.

Fig. 1: Surveyed district of West Kazakhstan Region.



Determination of the Therapeutic dose of Phytopreparation

For the research, we took the methodology for the determination of toxic impurities in flower pollen according to GOST 28887-90 «Flower pollen (beebread). Technical conditions». During the study, this method was adjusted to consider the work's purpose, the materials used, the drug's dosage form and the planned schemes and methods of treating bee colonies.

Approximately 60 bees were taken from one colony, extracted from a honeycomb frame with open brood, and placed in 15x6.5x13cm entomological cages. There were 2 bee cages in each group. The total number of bees in each group was at least 120.

The therapeutic dose of the phytopreparation was determined by administering the phytopreparation to bees in a dose-dependent manner. The experiment scheme included five groups of bees: four experimental and one control group.

For feeding bees in experimental groups, the tested preparation was mixed with 50% sugar syrup in volumes of 10, 15, 20 and 30 mL/1L of syrup. The syrup with the preparation was poured into test tubes. The test tubes for the control group were filled with 50% sugar syrup without preparation. All test tubes were covered with polyethylene film and secured with a rubber band. Small holes were made in the plastic film with a needle to allow the bees to take the food. The syrup tubes were inverted and placed in the cage, allowing the bees to access the food (Fig. 2). Bee cages were placed in a thermostat and kept at 30°C for 12 days (Fig. 3). The number of live and dead bees was counted daily. The average lifespan of bees in experimental and control groups was determined according to formula 1:

Po(k) = n1 + n2 + ... + n12 / N(1)

where Po(k) is the average lifespan of bees in experimental and control groups;

n1, n2, etc. - is the number of live bees in the cages on the corresponding day of counting;

N - the total number of bees in the group.

The average life expectancy of bees in experimental and control cages was the main criterion for assessing the harmlessness of the phytopreparation for bees. According to the methodology, the indicator in the experimental group should not be less than that in the control group. If the average lifespan of the experimental bees is shorter, the tested preparation or its concentration is considered toxic for bees.



Fig. 2: Bees in an entomological cage.



Fig. 3: Cages with bees in the thermostat.

Experimental Groups and Treatment Schemes for bee Colonies

For the scientific and farming experiment, 5 groups with 3 analog bee colonies in each group were formed during the spring period, specifically in the month of May. Before experimenting, a comprehensive assessment of the condition of bee colonies was carried out according to the following characteristics:

- infestation of bee colonies with varroosis (Varroa infestation is diagnosed based on visual detection of Varroa mites and laboratory data. (Methodological guidelines for the rapid diagnosis of varroosis and determination of the degree of infestation of bee colonies by Varroa mites in apiary conditions following methodological guidelines of the USSR Ministry of Agriculture (1984) - degree of nosemosis disease. To assess the influence of family strength on honey productivity, an indicator such as the relative amount of honey per 1kg of live bee weight is introduced. This indicator characterizes the ability of one kilogram of live bee weight (from the family under consideration) to collect a certain amount of honey. The viability (strength) of the bee colony was assessed before sampling and after treatment in accordance with the Instructions for the assessment (evaluation) of the breeding value and reproduction of bees, approved by order of the Minister of Rural Development of the Republic of Kazakhstan (2014). Moreover, assessed by the number of frames covered with bees on both sides in accordance with the requirements of GOST 20728-2014 Bee colony. Technical conditions.)

- the strength of bee colonies.

The mentioned groups of bee colonies were treated

with phytopreparation (phytopreparation in accordance with the recommended doses of preparations with a similar active substance. (Order No. 101 of March 6, 2018. On the approval of rules for conducting preclinical trials of medicinal products for veterinary use, clinical trials of medicinal products for veterinary use, and bioequivalence trials of medicinal products for veterinary use)) and known antivarroosis and antinociceptive preparations (as control) according to the scheme:

1. Phytopreparation method of feeding – 10 mL/1L of 50% sugar syrup 200 mL every other day for 12 days;

2. Phytopreparation method of feeding - 15mL/1L of 50% sugar syrup 200 mL every other day for 12 days;

3. Phytopreparation - irrigation - 10 mL/1L of 20% sugar syrup at the rate of 10-12 mL per 1 bee hive for 12 days every other day;

4. Phytopreparation - irrigation - 15 mL/1L of 20% sugar syrup at the rate of 10-12 mL per 1 bee hive for 12 days every other day;

5. Bipin-T+nosemat - according to instructions for use.

After treatment, the honey productivity of bee colonies was determined in the fall.

Sampling of Bees for Research

Sampling of bees to determine the infestation of bee colonies (intensity of infestation) was done from 15 bee colonies. Adult bees and bee dung collected near the hives were selected for the study. Each sample contained a minimum of 50 individuals for the study. Samples of bees were placed in an air-accessible plastic container and paper bags by shaking. Bee dung was collected separately in paper bags (Antonov, 1987).

Methods of Examination for Nosemosis

Microsporidia infestation of honey bees was analyzed according to «Methodological guidelines for laboratory tests for nosema of honey bees» (Antonov, 1987) using a microscope with an 8MP digital camera, LCD touch screen, and a hemocytometer. The number of spores of *Nosema spp.* was counted in the hemocytometer and used to calculate the average number of spores per bee.

Methods of Diagnosis of Varroosis

The diagnosis of varroosis was made after careful visual inspection of bee colonies and laboratory data. The studies were carried out in accordance with the approved in the territory of the Republic of Kazakhstan «Methodological guidelines for express-diagnosis of varroosis and determination of the degree of damage of bee colonies by varroa mites in apiary conditions».

A Method for Determining the Strength of bee Colonies

The viability (strength) of the bee colony was assessed before sampling and after treatment according to the Instruction on Bonding (Evaluation) of Breeding Value and Reproduction of Bees, approved by the Minister of Rural Development of the Republic of Kazakhstan (2014). Moreover, evaluated by the number of honeycombs sitting on both sides of the bees by the requirements of GOST 20728-2014 «Bee colony. Technical conditions».

Method for Determining Honey Productivity

The honey productivity of the apiary was estimated by the total amount of honey produced. For this purpose, the amount of marketable honey and the amount of forage honey remaining in the hive were counted. The amount of honey in the honeycomb was determined by one of the following methods: weighing the frames by subtracting the weight of the empty honeycomb on a scale, or visually by measuring the area of the honeycomb filled with honey. It was considered that a fully sealed honeycomb frame with a volume of 43.5×30.0 cm up to 3kg, and 43.5×23.0 cm up to 3kg would be suitable. The amount of sugar fed at the beginning of the season was subtracted from the amount of gross honey (Seriakov, 2021).

Statistical Method

Accumulation, correction, systematization of initial information, statistical processing, and analysis of results were performed using Microsoft Office Excel 2016 spreadsheets. Statistical analysis was also performed using online calculators https://medstatistic.ru (Marapov, 2022).

The results and the difference between treatment options were analyzed using Student's t-test. Data are presented as mean+SE. For this purpose, the following main parameters were calculated: mean, variance, standard deviation, standard error, and significance level. The significance level to determine the statistical significance of the observed differences between treatment options was P < 0.05.

RESULTS

At the first stage of the study, the infestations of nosema and varroa in bee colonies were assessed. Based on clinical and laboratory tests, nosema (Fig. 4) and varroa (Fig. 5) infestations were identified.

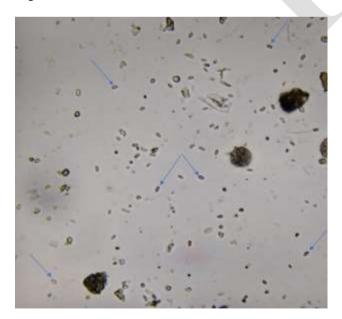


Fig. 4: Nosema spores at 400x magnification.

To study the therapeutic efficacy of phytopreparation, experimental and control groups of bee colonies were formed according to the principle of analog colonies.



Fig. 5: Varroa mite.

Experimenting to Determine the Harmlessness of the Phytopreparation for bees and to Establish its Therapeutic Dose

To establish the therapeutic dose, the phytopreparation was tested by feeding method at the dose of 10, 15, 20 and 30 mL of phytopreparation per 1L of sugar syrup.

The results of the experiment are presented in Table 1. As can be seen from Table 1, the average life expectancy of bees in the control cage was 10.55 days, according to the results. The average life expectancy of bees receiving phytopreparation in different dilutions differs from each other and the control indicator. When using the phytopreparation in the dose of 10-15mL/1L of sugar syrup, the average life expectancy of bees exceeds the control indicator, and according to the methodology, these doses are harmless to bees. They can be used for testing on bee colonies. Using the phytopreparation in supplementary feeding at dilutions of 20 and 30 mL of the phytopreparation per 1L of sugar syrup causes poisoning and death of bees. Thus, these dilutions can be considered toxic for bees. They are excluded from further experiments. The following doses are recommended for scientific and economic experiments: 10 and 15 mL of the preparation per 1L per second.

Table	1: Ave	rage	lifespan	of Ł	bees in	control	and	experi	imental	cage	5
	-										

Nº	Preparation	Average lifespan
		of bees, days
1	Control	10.55
2	Phytopreparation 10mL/1L of sugar syrup	11.28
3	Phytopreparation 15mL/1L of sugar syrup	11.62
4	Phytopreparation 20mL/1L liter of sugar syrup	10.54
5	Phytopreparation 30mL/1L of sugar syrup	10.33

Results of a Scientific and Economic Experiment

After spring treatment with the phytopreparation, a general excitement of bees was observed and their cleansing and summer activity was activated.

As shown in Table 2 (Fig. 6), the stickiness of bee colonies before treatment averaged 7.8-8.0% in the groups. As a result of treatments with phytopreparation by feeding

Table 2: Effectiveness of phytopreparation in spring treatment of bee colonies	tion in spring tr	eatment of be	ee colonies										
	Alleyw	Alleyways, pcs.	Frames,	quantity	Frames with brood,	th brood,	Honey productivity,		Varroosis incidence, %	ice, %	Nosema in	festation, nun	Nosema infestation, number of spores
					number	iber	gross honey, kg					per 1 bee	
	Before	After	Before	After	Before	After		Before	After	Decrease	in Before	After	Decrease in
	treatment	treatment	treatment	treatment	treatment	treatment		treatment	treatment	lesions, %	treatment	treatment	lesions, %
1. Phytopreparation method	of 8	18	16	20	5	14	52	8.1	1.3	84			
feeding – 10 mL/1L of 50% sugar syrup	C 7	18	16	19	9	12	45	7.5	0.9	88			
	7	18	16	20	J	12	45	7.8	1.1	86			
Average	7.3±0.4	18.0 ± 0.0	16±0.0	19.7±0.4	5.3±0.4	12.7±1.2	47.3±2.9	7.8 ±0.2	1.1±0.1	86			
2. Phytopreparation method	of 8	18	16	20	5	13	45	8.3	1.0	88	12		100
feeding - 15mL/1L of 50% sugar syrup	8	18	16	20	5	13	50	7.6	1.1	85	15		100
	7	17	15	19	5	12	43	7.4	1.2	84	9		100
Average	7.7±0.4	17.7±0.4	15.7±0.4	19.7±0.4	5.0±0.4	12.7±0.4	46.0±2.5	7.7±0.3	1.1±0.16	85.7	11±3.2		100
3. Phytopreparation - irrigation - 10 8	10 8	18	16	20	5	12	47	7.6		84	ı	,	
mL/1L of 20% sugar syrup	7	18	16	20	5	13	49	8.0	1.2	84			
	8	18	16	20	9	12	44	7.9	1.2	85	ı		
Average	7.7±0.4	18.0 ± 0.0	16.0 ± 0.0	20±0	5.3±0.4	12.3±0.4	46.7±1.8	7.8±0.15	1.2 ± 0.05	84.3	ı		
4. Phytopreparation - irrigation - 15	15 7	17	16	20	9	13	49	7.9	1.3	83	'	,	
mL/1L of 20% sugar syrup	8	18	15	20	5	12	42	8.1	1.1	86	ı	,	
Average	7.7±0.4	17.7±0.4	15.7±0.4	20.0±0.0	5.3±0.4	12.3±0.4	45.3±2.5	8.0±0.07	1.2 ± 0.05	85	ı	,	
Bipin-T + nosemat	7	17	16	20	5	12	45	7.5	1.4	81	7	,	100
	8	18	16	19	6	12	44	7.9	1.2	85	13	,	100
	8	18	16	19	5	12	43	8.2	1.2	85	8	,	100
Average	7.7±0.4	17.7±0.4	16.0±0.0	19.3±0.4	5.3±0.4	12.0±0.0	44.0±0.7	7.9±0.23	1.3±0.1	83.5	9.3±2.3	ı	100

at doses of 10 and 15 mL/1L of sugar syrup, the stickiness of bee colonies in both groups decreased to 1.1 % or by 86.0 and 85.7%, respectively. At irrigation in the same doses, the therapeutic efficacy of the phytopreparation amounted to 84.3 and 85%. The use of Bipin-T reduced the pecking rate by 83.5%.

In the bee colonies of the second and fifth groups before treatment, a weak degree of nosemosis (ranging from 1 to 15 spores in the microscope field of view) was observed; in the rest, no disease was detected. After treatment, all groups of bee colonies were found to be healthy for nosomosis.

When determining the statistical significance of the observed differences between treatment options, the significance level was above the established level (P> 0.05) in all cases and for all considered indicators. Consequently, the phytopreparation corresponds to such preparations as Bipin-T and nosemat in terms of therapeutic efficacy.

Additionally, in the groups treated with phytopreparation, the number of brood frames was slightly higher than in the control group. However, this difference is not significant (P>0.05).

DISCUSSION

The use of medicinal plants in beekeeping for the prevention and treatment of bee diseases or as biologically active food additives gives a comprehensive positive effect, manifested in increased survival rate of bees, reduction in the number or complete elimination of the causative agent of the disease, reduction of stress in bees (Sofou et al., 2017; Jovanovic et al., 2023). Plant preparations may not have a significant effect on bee immunity (Glavinic et al., 2024). However, when used, bee mortality is significantly lower than when treated with chemical preparations (Rahimi & Prichrhreh, 2024).

The positive effects of medicinal plants and their preparations are observed when used on sick bee colonies. However, there is an opinion that in healthy bees, some plants and their active substances can cause certain disturbances in the bee organism, reducing their survival rate and immunity (Potrich et al., 2020; Glavinic et al., 2022).

Consequently, when using preparations for disease prevention, it is essential to exercise caution and use only proven remedies and plants with a proven effect on the bee organism.

For this purpose, laboratory tests are conducted to determine the therapeutic dose of herbal preparations, thereby establishing optimal schemes and methods of treatment for bee colonies (Ariana et al., 2002; Sofou et al., 2017; Vilarem et al., 2021). In our study, the laboratory experiment showed that phytopreparation is safe for bees at doses of 10 and 15 mL/1L of sugar syrup.

It should be noted that when using alcoholic extracts of plants, it is necessary to take into account the concentration of alcohol in sugar syrup, as ethanol at a concentration of more than 2.5% in complex with the pathogen can cause a synergistic effect and significantly reduce the viability of

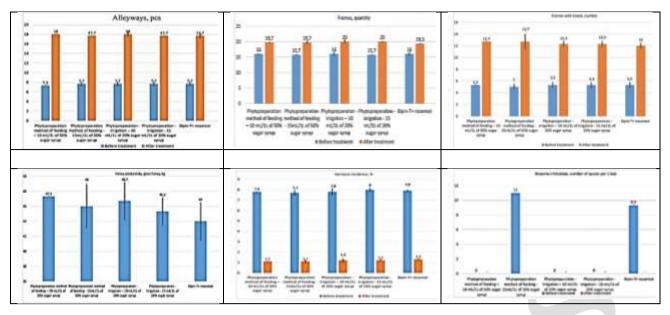


Fig. 6: Effectiveness of phytopreparation in spring treatment of bee colonies.

bees (Ptaszyńska et al., 2013). The dilutions of the phytopreparation (10 and 15 mL/1L of syrup) comply with these requirements and do not exceed the established concentration of ethanol in the working solution.

It is believed that the therapeutic dose of the preparation and its efficacy depend on the route of administration (Ozkırım et al., 2021; Castagna et al., 2022; Bava et al., 2023).

According to available data, one of the most effective ways to use plant preparations is to feed with a 50% sucrose solution containing 0.1-5% plant extract. In this case, their effectiveness is comparable to known chemotherapeutic preparations (Allabergenova et al., 2021).

The acaricidal and insecticidal activity of spraying alcoholic extracts of plants also has a high level of toxicity against mites and is harmless to bees (Damiani et al., 2011). The therapeutic efficacy can reach 100% in some cases (Razaviet al., 2015; Allabergenova et al., 2021).

The results of our studies confirm this. In the treatment of varroosis, the therapeutic efficacy of the phytopreparation at feeding was 86 and 85.7% and at irrigation, 84.3 and 85%. In the control, mite infestation decreased by 83.5%. In the treatment of nosemosis, the efficacy of the phytopreparation and control was 100%.

The results of these studies, as well as the high efficacy of plant alcohol extracts, are confirmed by the studies of other scientists. For example, when studying 90% ethanol extracts of leaves and stem bark of *Swietenia mahogani* and *Swietenia macrophylla*, acaricidal activity reached 100% and 95%, respectively (El Zalabani et al., 2012). The ethanol extract of laurel leaves, after a 30-second exposure, kills 50% of mites within 24 hours after treatment (Damiani et al., 2014). The use of a mixture of extracts from T. thymus and *H. perforatum* in Kazakhstan resulted in a reduction of *Varroa* mite abundance index to 0% (Allabergenova et al., 2021).

In the treatment and prevention of nosemosis, the effectiveness of phytopreparations is also high. In bee colonies that received plant extracts with feeding, fewer dead bees and lower levels of nosema infestation were

observed during the winter period (Ptaszyńska et al., 2013). This may be due to the fact that plant extracts at a concentration of 1-2% inhibit the development of *Nosema* spores already on the 5th-9th day after treatment, the number of microsporidium spores decreases 4-6 times within 30 days after treatment and the efficacy is 32.5-100% (Damiani et al., 2014; Chaimanee et al., 2021; El-Sayed et al., 2024). The antimicrobial activity of the plant extract is mainly due to the presence of phenolic compounds and terpenoids (lorizzo et al., 2022).

The use of phytopreparation, in most cases, also has a bio stimulating effect on bees. In our study, after treatment, the number of brood frames in the experimental groups was slightly higher than in the control group; however, the difference was not significant (P>0.05). In all groups treated with phytopreparation, honey production of bee colonies was also higher than in the control. Although the significance level of the observed differences between the treatment variants exceeds the established value (P>0.05), this suggests that the studied phytopreparation has not only a therapeutic effect but also favorably affects all economically valuable traits of bee colonies. This may be because the addition of phytopreparation to sugar syrup increases the amount of carbohydrates and protein in the bee body, increases the productivity of bee colonies (Amera et al., 2024) and as a consequence, increases the honey productivity of bee colonies and improves the quality of honey (Mahmood et al., 2014; Vasileva et al., 2024).

Thus, the results of this study enable us to recommend phytopreparation for use in veterinary practice as an effective means of preventing and treating varroosis and nosemosis in bees, thereby increasing the natural resistance of bee colonies to diseases and enhancing their productivity.

Phytopreparation has several advantages: it has a pronounced antivarroosis, antinosemosis, and biostimulating effect. The phytopreparation has no side effects during its use. The components of the phytopreparation are environmentally friendly, readily available in the Republic of Kazakhstan and cost-effective. The method of phytopreparation manufacturing is laborintensive and low-cost.

The appearance of highly effective, inexpensive, and non-toxic to bees and humans therapeutic and prophylactic preparations from plant raw materials on the market will improve the epizootic situation in apiaries, allowing for the preservation and increase in the number of bee colonies and the production of high-quality industrial products.

Conclusion

The results of the present study showed that the investigated phytopreparation used in the spring period by the method of feeding at a dose of 10-15 mL/1L of 50% sugar syrup or by irrigation at the same dose in 20% sucrose syrup is a highly effective environmentally friendly preparation that can be successfully used in the practice of beekeeping for the prevention and treatment of varroosis and nosemosis of bees, to increase the natural resistance of bee colonies to diseases, to increase their productivity. The practical use and results of this study demonstrate the feasibility of phytopreparations for preventing and treating varroosis and nosemosis in bees as an effective, safe, and affordable means.

It is essential to continue studying the phytopreparation on a larger number of experimental bee colonies to determine its effectiveness in the fall treatment of bee colonies, as well as to investigate its impact on the organism and economically valuable characteristics of bees.

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Author's Contribution: BS, FN, NV, and GK collected data, implemented experiments, conducted experimental work, and analyzed and interpreted data. NV developed the concept and content of the research, as well as writing and designing the article. KK supervised the research, collated the research results, reviewed, and edited the article. SB conducted statistical analysis and interpretation of results. AK and EM prepare literature selection, compilation of bibliographies, and analysis of processed information. AK prepares draft articles, including template frameworks and modifications to the final submitted version. All authors read and approved the final version of the manuscript.

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REFERENCES

- Acting Minister of Ecology and Natural Resources of the Republic of Kazakhstan (2023). Rules for the management of vegetation. Order of the Acting Minister of Ecology and Natural Resources of the Republic of Kazakhstan No 137 of 28 April 2023. Registered with the Ministry of Justice of the Republic of Kazakhstan on 3 May 2023 No 32415.
- Ahmad, D., & Elsaiegh, M. (2022). The use of different concentrations of oxalic acid to decrease the infestation of Varroa destructor during autumn season as one of integrated management and methods. NTU Journal of Agriculture and Veterinary Science, 2(2), 30-37. https://doi.org/10.56286/ntujavs.v2i2.294
- Allabergenova, A., Turganbayeva, G., & Nurseitova, M. (2021). Control of Varroa destructor in Kazakhstan. Archives of Razi Institute, 76(5), 1389– 1397. <u>https://doi.org/10.22092/ari.2021.355621.1704</u>
- Amera, W.A., Mersso, B.T., Sisay, T.A., Arega, A.B., & Alene, A.T. (2024). Effect of various supplements on productive performance of honey bees, in the South Wollo Zone, Ethiopia. *PloS One*, 19(5): e0303579. <u>https://doi.org/10.1371/journal.pone.0303579</u>
- Antonov, B.I. (1987). Laboratory investigations in veterinary medicine: viral, rickettsial and parasitic diseases. Moscow: Agropromizdat, 240.
- Ariana, A., Ebadi, R., & Tahmasebi, G. (2002). Laboratory evaluation of some plant essences to control Varroa destructor (Acari: Varroidae). *Experimental Applied Acarology*, 27(4): 319–327. <u>https://doi.org/10.1023/a:1023342118549</u>
- Aurell, D., Wall, C., Bruckner, S., & Williams, G.R. (2024). Combined treatment with amitraz and thymol to manage Varroa destructor mites (Acari: Varroidae) in Apis mellifera honey bee colonies (Hymenoptera: Apidae). Journal of Insect Science, 24(3): 12. https://doi.org/10.1093/jisesa/ieae022
- Baigazanov, A., Tikhomirova, Y., Valitova, N., Nurkenova, M., Koigeldinova, A., Abdullina, E., Zaikovskaya, O., Ikimbayeva, N., Zainettinova, D., & Bauzhanova, L. (2022). Occurrence of nosemosis in honey bee, *Apis* mellifera L. At the apiaries of East Kazakhstan. PeerJ, 10: e14430. https://doi.org/10.7717/peerj.14430
- Bava, R., Castagna, F., Lupia, C., Ruga, S., Musella, V., Conforti, F., Marrelli, M., Argentieri, M.P., Britti, D., Statti, G., & Palma, E. (2023). Chemical profile of essential oils of selected *Lamiaceae* plants and in vitro activity for varroosis control in honeybees (*Apis mellifera*). *Veterinary Sciences*, 10(12), 701. https://doi.org/10.3390/vetsci10120701
- Bava, R., Castagna, F., Palma, E., Ceniti, C., Millea, M., Lupia, C., Britti, D., & Musella, V. (2023). Prevalence of *Varroa destructor* in honeybee (*Apis mellifera*) farms and varroosis control practices in Southern Italy. *Microorganisms*, 11(5), 1228. <u>https://doi.org/10.3390/microorganisms11051228</u>

- Bava, R., Castagna, F., Palma, E., Marrelli, M., Conforti, F., Musolino, V., Carresi, C., Lupia, C., Ceniti, C., Tilocca, B., Roncada, P., Britti, D., & Musella, V. (2023). Essential oils for a sustainable control of honeybee varroosis. *Veterinary Sciences*, 10(5), 308. <u>https://doi.org/10.3390/vetsci10050308</u>
- Bava, R., Castagna, F., Ruga, S., Nucera, S., Caminiti, R., Serra, M., Bulotta, R. M., Lupia, C., Marrelli, M., Conforti, F., Statti, G., Domenico, B., & Palma, E. (2023). Plants and their derivatives as promising therapeutics for sustainable control of honeybee (*Apis mellifera*) pathogens. *Pathogens (Basel, Switzerland)*, 12(10), 1260. https://doi.org/10.3390/pathogens12101260
- Bekele, A.Z., Mor, S.K., Phelps, N.B.D., Goyal, S.M., & Armién, A.G. (2015). A case report of *Nosema ceranae* infection in honey bees in Minnesota, USA. Veterinary Quarterly, 35(1), 48–50. https://doi.org/10.1080/01652176.2014.981766
- Boonmee, T., Sinpoo, C., Wongthaveethong, L., Disayathanoowat, T., Suanpoot, P., Pettis, J.S., & Chaimanee, V. (2024). Properties of essential oils absorbed on the surface of cardboard pieces after using atmospheric-pressure plasma treatments to develop long-lasting *Varroa* miticides in honeybees (*Apis mellifera*). *PLoS One*, 19(2), e0297980. <u>https://doi.org/10.1371/journal.pone.0297980</u>
- Castagna, F., Bava, R., Piras, C., Carresi, C., Musolino, V., Lupia, C., Marrelli, M., Conforti, F., Palma, E., Britti, D., & Musella, V. (2022). Green veterinary pharmacology for honey bee welfare and health: Origanum heracleoticum L. (Lamiaceae) essential oil for the control of the Apis mellifera varroatosis. Veterinary Sciences, 9(3), 124. https://doi.org/10.3390/vetsci9030124
- Chaimanee, V., Kasem, A., Nuanjohn, T., Boonmee, T., Siangsuepchart, A., Malaithong, W., Sinpoo, C., Disayathanoowat, T., & Pettis, J.S. (2021). Natural extracts as potential control agents for *Nosema ceranae* infection in honeybees, *Apis mellifera. Journal of Invertebrate Pathology*, 186: 107688. <u>https://doi.org/10.1016/j.jip.2021.107688</u>
- Chemurot, M., Akol, A.M., Masembe, C., de Smet, L., Descamps, T., & de Graaf, D.C. (2016). Factors influencing the prevalence and infestation levels of *Varroa destructor* in honeybee colonies in two highland agro-ecological zones of Uganda. *Experimental Applied Acarology*, 68(4), 497–508. <u>https://doi.org/10.1007/s10493-016-0013-x</u>
- Cilia, G., & Nanetti, A. (2023). Challenges and advances in bee health and diseases. *Veterinary Sciences*, 10(4), 253. https://doi.org/10.3390/vetsci10040253
- Council of the Eurasian Economic Commission (2018). On the introduction of the Rules of the above-mentioned practice of the manning, collection, processing and preservation of the eligible cheese of the growing procuring entity. Resolution of the Council of the Eurasian Economic Commission No 15 of 26.01.2018. https://adilet.zan.kz/rus/docs/H18EV000015
- Damiani, N., Fernández, N.J., Porrini, M.P., Gende, L.B., Álvarez, E., Buffa, F., Brasesco, C., Maggi, M.D., Marcangeli, J.A., & Eguaras, M.J. (2014). Laurel leaf extracts for honeybee pest and disease management: antimicrobial, microsporicidal, and acaricidal activity. *Parasitology Research*, 113(2), 701–709. <u>https://doi.org/10.1007/s00436-013-3698-3</u>
- Damiani, N., Gende, L.B., Maggi, M.D., Palacios, S., Marcangeli, J.A., & Eguaras, M.J. (2011). Repellent and acaricidal effects of botanical extracts on Varroa destructor. Parasitology Research, 108(1), 79–86. https://doi.org/10.1007/s00436-010-2043-3
- Dequenne, I., Philippart de Foy, J.M., & Cani, P.D. (2022). Developing strategies to help bee colony resilience in changing environments. *Animals*, 12(23), 3396. https://doi.org/10.3390/ani12233396
- El Zalabani, S.M., El-Askary, H.I., Mousa, O.M., Issa, M.Y., Zaitoun, A.A., & Abdel-Sattar, E. (2012). Acaricidal activity of Swietenia mahogani and Swietenia macrophylla ethanolic extracts against Varroa destructor in honeybee colonies. Experimental Parasitology, 130(2), 166–170. https://doi.org/10.1016/j.exppara.2011.10.013
- El-Sayed, A.S.A., Fathy, N.A.M., Labib, M., El-Baz, A.F., El-Sheikh, A.A., & Moustafa, A.H. (2024). Biological control of nosemosis in *Apis mellifera* L. with *Acacia nilotica* extract. *Scientific Reports*, 14(1), 28340. <u>https://doi.org/10.1038/s41598-024-78874-6</u>
- El-Seedi, H.R., El-Wahed, A.A.A., Naggar, Y.A., Saeed, A., Xiao, J., Ullah, H., Musharraf, S.G., Boskabady, M.H., Cao, W., Guo, Z., Daglia, M., El Wakil, A., Wang, K., & Khalifa, S.A.M. (2022). Insights into the role of natural products in the control of the honey bee gut parasite (*Nosema spp.*). *Animals*, 12(21), 3062. <u>https://doi.org/10.3390/ani12213062</u>
- Er Demirhan, B., & Demirhan, B. (2022). Detection of antibiotic residues in blossom honeys from different regions in Turkey by LC-MS/MS method. *Antibiotics*, 11(3), 357. <u>https://doi.org/10.3390/antibiotics11030357</u>
- Eurasian Economic Commission (2023). Recommendation of the Eurasian Economic Commission of 14.11.2023 №33 "On the Guide for the use of laboratory (experimental) animals in the conduct of non-clinical

research". https://online.zakon.kz/Document/?doc_id=37697592 (date of circulation 03.03.2025).

- Farida, N., Satybaev, B., Kairgalieva, G., Kushaliev, K., Baiantasova, S., Valitova, N.V., Kozhaeva, A., & Agisheva, E.R. (2024). Epizootologicheskii monitoring v pchelokhoziaistvakh Zapadno-Kazakhstanskoi oblasti. *Nauka i Obrazovanie*, 1(3), 163–173. <u>https://doi.org/10.52578/2305-9397-2024-3-1-163-173</u>
- Formato, G., Rivera-Gomis, J., Bubnic, J., Martín-Hernández, R., Milito, M., Croppi, S., & Higes, M. (2022). Nosemosis prevention and control. *Applied Sciences*, 12(2), 783. <u>https://doi.org/10.3390/app12020783</u>
- Formato, G., & Smulders, F.J.M. (2011). Risk management in primary apicultural production. Part 1: bee health and disease prevention and associated best practices. *Veterinary Quarterly*, 31(1), 29–47. <u>https://doi.org/10.1080/01652176.2011.565913</u>
- Formato, G., Zilli, R., Condoleo, R., Marozzi, S., Davis, I., & Smulders, F.J.M. (2011). Risk management in primary apicultural production. Part 2: a Hazard Analysis Critical Control Point approach to assuring the safety of unprocessed honey. *Veterinary Quarterly*, 31(2), 87–97. <u>https://doi.org/10.1080/01652176.2011.567755</u>
- Gilbert, J., Paris, L., Dubuffet, A., Texier, C., Delbac, F., & Diogon, M. (2024). Nosema ceranae infection reduces the fat body lipid reserves in the honeybee Apis mellifera. Journal of Invertebrate Pathology, 207, 108218. https://doi.org/10.1016/j.jip.2024.108218
- Glavinic, U., Blagojevic, J., Ristanic, M., Stevanovic, J., Lakic, N., Mirilovic, M., & Stanimirovic, Z. (2022). Use of thymol in *Nosema ceranae* control and health improvement of infected honey bees. *Insects*, 13(7), 574. <u>https://doi.org/10.3390/insects13070574</u>
- Glavinic, U., Jovanovic, N.M., Dominikovic, N., Lakic, N., Ćosić, M., Stevanovic, J., & Stanimirovic, Z. (2024). Potential of wormwood and oak barkbased supplement in health improvement of *Nosema ceranae*-infected honey bees. *Animals*, 14(8), 1195. <u>https://doi.org/10.3390/ani14081195</u>
- GOST 20728-2014. Bee family. Technical conditions. Substitute GOST 20728-75; vved. (2016-07-01). Moskva: Standartinform: 6
- GOST 28887-90. Flower pollen (obnozhka). Technical conditions. Group S52. MKS 65.140. OKSTU 9882. Date of entry (1991-07-01).
- Grobov, O.F., Sotnikov, A.N., & Shtondina, D.A. (2008). Relationship of *Varroa* destructor with various organisms. *Veterinary Pathology*, 3, 5.
- Guün, R., Dursun, I., Arıcı, B., & Sarac, Y. (2024). Detection of multiple antibiotic residues in Turkish pine and blossom honeys using LC-MS/MS method. *Chemistry and Biodiversity*, 21(1), e202301261. https://doi.org/10.1002/cbdv.202301261
- Iorizzo, M., Letizia, F., Ganassi, S., Testa, B., Petrarca, S., Albanese, G., Di Criscio, D., & De Cristofaro, A. (2022). Recent advances in the biocontrol of nosemosis in honey bees (*Apis mellifera* L.). *Journal of Fungi*, 8(5), 424. <u>https://doi.org/10.3390/jof8050424</u>
- Jack, C.J., Boncristiani, H., Prouty, C., Schmehl, D.R., & Ellis, J.D. (2024). Evaluating the seasonal efficacy of commonly used chemical treatments on Varroa destructor (Mesostigmata: Varroidae) population resurgence in honey bee colonies. Journal of Insect Science, 24(3), 11. https://doi.org/10.1093/jisesa/ieae011
- Jovanovic, N.M., Glavinic, U., Ristanic, M., Vejnovic, B., Ilic, T., Stevanovic, J., & Stanimirovic, Z. (2023). Effects of plant-based supplement on oxidative stress of honey bees (*Apis mellifera*) infected with *Nosema ceranae*. *Animals*, 13(22), 3543. <u>https://doi.org/10.3390/ani13223543</u>
- Kerimaliev, Z.K. (2005). Invasive diseases of honey bees in Kyrgyzstan and prospects for their elimination. Proceedings of the International Scientific and Practical Conference "Current state and actual problems of development of veterinary science and practice" dedicated to the 100th anniversary of KazNIVI, T.2, Invazionnye i nezaraznye bolezni, Almaty, 83-86.
- Khan, S.U., Anjum, S.I., Ansari, M.J., Khan, M.H., Kamal, S., Rahman, K., Shoaib, M., Man, S., Khan, A.J., Khan, S.U., & Khan, D. (2019). Antimicrobial potentials of medicinal plant extracts and their derived silver nanoparticles: A focus on honey bee pathogens. *Saudi Journal of Biological Sciences*, 26(7), 1815–1834. https://doi.org/10.1016/j.sjbs.2018.02.010.
- Kunat-Budzynńska, M., Budzynński, M., Schulz, M., Strachecka, A., Gancarz, M., Rusinek, R., & Ptaszynńska, A.A. (2022). Natural substances, probiotics, and synthetic agents in the treatment and prevention of honeybee nosemosis. *Pathogens*, 11(11), 1269. <u>https://doi.org/10.3390/pathogens11111269</u>
- Lambert, O., Piroux, M., Puyo, S., Thorin, C., L'Hostis, M., Wiest, L., Buleteé, A., Delbac, F., & Pouliquen, H. (2013). Widespread occurrence of chemical residues in beehive matrices from apiaries located in different landscapes of Western France. *PLoS One*, 8(6), e67007. <u>https://doi.org/10.1371/journal.pone.0067007</u>
- Larkina, E.O., & Lapynina, E.P. (2021). Effect of antivarroatosis preparations on external characteristics of honey bees. *Bulletin of Agrarian Science*,

6(93), 32-36.

- Lozano, A., Hernando, M.D., Uclés, S., Hakme, E., & Fernández-Alba, A.R. (2019). Identification and measurement of veterinary drug residues in beehive products. *Food Chemistry*, 274, 61–70. <u>https://doi.org/10.1016/j.foodchem.2018.08.055</u>
- Mahmood, R., Asad, S., Raja, S., Atta-ul-Moshin, Wagchoure, E.S., Sarwar, G. R., Islam, N., & Ahmad, W. (2014). Control of Varroa destructor (Acari: Varroidae) in Apis mellifera (Hymenoptera: Apidae) by using plant oils and extracts. *Pakistan Journal of Zoology*, 46, 609-615.
- Marapov, D. (2022). Medical Statistics. Electronic resource. Access mode: https://medstatistic.ru.
- Mayack, C., & Naug, D. (2009). Energetic stress in the honeybee Apis mellifera from Nosema ceranae infection. Journal of Invertebrate Pathology, 100(3), 185–188. <u>https://doi.org/10.1016/j.jip.2008.12.001</u>
- Mikhaltsevich, G.N., & Velichko, M.G. (1996). Pathogenic flora on the background of varroatosis. *Beekeeping*, 1, 26-28.
- Minister of Rural Development of the Republic of Kazakhstan (2014). Order of the Minister of Rural Development of the Republic of Kazakhstan No 277 of 28 July 2023. Registered with the Ministry of Justice of the Republic of Kazakhstan on 31 July 2023 under No 33191. On introducing changes and amendments to the Order of the Minister of Rural Development of the Republic of Kazakhstan of 10 October 2014 No 3-3/517 "On the implementation of the instruction on the evaluation of the breeding status and breeding of livestock". https://adilet.zan.kz/rus/docs/V2300033191#z6 (date of publication 25.01.2025).
- Noël, A., Le Conte, Y., & Mondet, F. (2020). Varroa destructor: how does it harm Apis mellifera honey bees and what can be done about it? *Emerging Topics in Life Sciences*, 4(1), 45-57. <u>https://doi.org/10.1042/ETLS20190125</u>
- Ostap-Chec, M., Cait, J., Scott, R.W., Arct, A., Moroń, D., Rapacz, M., & Miler, K. (2024). Nosemosis negatively affects honeybee survival: experimental and meta-analytic evidence. *Parasitology*, 151(14), 1530-1542. https://doi.org/10.1017/S0031182024001446
- Ozkırım, A., & Kucukozmen, B. (2021). Application of herbal essential oil extract mixture for honey bees (Apis mellifera L.) against Nosema ceranae and Nosema apis. *Journal of Apicultural Science*, 65, 163-175. <u>https://doi.org/10.2478/jas-2021-0010</u>
- Ozuicli, M., Girisgin, A., Diker, A., Baykalır, Y., Kisadere, I., & Aydin, L. (2023). The efficacy of thyme, peppermint, eucalyptus essential oils, and nanoparticle ozone on nosemosis in honey bees. *Kafkas Universitesi Veteriner Fakultesi Dergisi*, 29(4), 335-342. https://doi.org/10.9775/kvfd.2023.29167
- Panek, J., Paris, L., Roriz, D., Mone, A., Dubuffet, A., Delbac, F., Diogon, M., & El Alaoui, H. (2018). Impact of the microsporidian Nosema ceranae on the gut epithelium renewal of the honeybee, Apis mellifera. *Journal of Invertebrate Pathology*, 159, 121–128. <u>https://doi.org/10.1016/j.jip.2018.09.007</u>
- Pasca, C., Matei, I.A., Diaconeasa, Z., Rotaru, A., Erler, S., & Dezmirean, D.S. (2021). Biologically active extracts from different medicinal plants tested as potential additives against bee pathogens. *Antibiotics*, 10(8), 960. <u>https://doi.org/10.3390/antibiotics10080960</u>
- Potrich, M., Silva, R.T., Maciel, R.M., Costa-Maia, F.M., Lozano, E.R., Rossi, R. M., Martins, J.R., & Dallacort, S. (2020). Are plant extracts safe for honey bees (Apis mellifera)? *Journal of Apicultural Research*, 59, 844-851.
- Ptaszyńska, A.A., Borsuk, G., Mułenko, W., & Olszewski, K. (2013). Impact of ethanol on Nosema spp. infected bees. *Medycyna Weterynaryjna*, 69(12), 736-740.
- Rahimi, A., & Prichrhreh, S. (2024). Evaluation of a new plant-based formulation to control Varroa mite (Varroa destructor) in honey bee (Apis mellifera) colonies. *Journal of Entomological Society of Iran*, 44(4), 417-428. https://doi.org/10.61186/jesi.44.4.5
- Rashid, B., Khani, A., Ghasemi, V., Ghadamyari, M., Sahebzadeh, N., & Moharramipour, S. (2020). Evaluation of a new plant-based formulation for the treatment of varroosis in the honey bee colonies: efficacy and safety. *Apidologie*, 51, 1074-1090. <u>https://doi.org/10.1007/s13592-020-00786-x</u>

- Razavi, S.M., Asadpour, M., Jafari, A., & Malekpour, S.H. (2015). The field efficacy of Lepidium latifolium and Zataria multiflora methanolic extracts against *Varroa destructor*. *Parasitology Research*, 114(11), 4233–4238. https://doi.org/10.1007/s00436-015-4661-2
- Reams, T., & Rangel, J. (2022). Understanding the enemy: A review of the genetics, behavior and chemical ecology of *Varroa destructor*, the parasitic mite of *Apis mellifera*. *Journal of Insect Science*, 22(1), 18. <u>https://doi.org/10.1093/jisesa/ieab101</u>
- Robi, D.T., Temteme, S., Aleme, M., Bogale, A., Getachew, A., & Mendesil, E. (2023). Epidemiology, factors influencing prevalence and level of varroosis infestation (*Varroa destructor*) in honeybee (*Apis mellifera*) colonies in different agroecologies of Southwest Ethiopia. *Parasite Epidemiology and Control*, 23, e00325. <u>https://doi.org/10.1016/j.parepi.2023.e00325</u>
- Rodrigues, H., Leite, M., Oliveira, B., & Freitas, A. (2024). Antibiotics in honey: a comprehensive review on occurrence and analytical methodologies. *Open Research Europe*, 4, 125. https://doi.org/10.12688/openreseurope.17664.2
- Salkova, D., Gurgulova, K., & Zhelyazkova, I. (2024). Clinical trial of the efficiency of three different compositions of acaricidal substances against varoosis in honey bee colonies. *Ankara Üniversitesi Veteriner Fakültesi Dergisi*, 71(2), 157-163. https://doi.org/10.33988/auvfd.1033097
- Seriakov, I.S. (2021). Beekeeping. Tribal work in beekeeping: a teaching and methodological manual. *Gorki: BGSKhA*, 60.
- Shakarian, G.A. (1981). Obtaining antibiotics in the body of bees. *Beekeeping*, 1, 21-23.
- Shoaei, F., Talebi-Ghane, E., Ranjbar, A., & Mehri, F. (2024). Evaluation of antibiotic residues in honey: a systematic review and meta-analysis. *International Journal of Environmental Health Research*, 34(2), 1064– 1075. <u>https://doi.org/10.1080/09603123.2023.2197285</u>
- Sofou, K., Isaakidis, D., Spyros, A., Büttner, A., Giannis, A., & Katerinopoulos, H.E. (2017). Use of costic acid, a natural extract from *Dittrichia viscosa*, for the control of *Varroa destructor*, a parasite of the European honey bee. *Beilstein Journal of Organic Chemistry*, 13, 952–959. https://doi.org/10.3762/bjoc.13.96
- Sotnikov, A.N. (1982). Acaricidal activity and toxicity of some preparations after feeding to bees with varroatosis. *Biul. VIEV*, 41, 70-72.
- Terpin, B., Perkins, D., Richter, S., Leavey, J.K., Snel, T.W., & Pierson, J.A. (2019). A scientific note on the effect of oxalic acid on honey bee larvae. *Apidologie*, 50, 363–368. <u>https://doi.org/10.1007/s13592-019-00650-7</u>
- Tokarev, Y.S., Zinatullina, Z.Y., Ignatieva, A.N., Zhigileva, O.N., Malysh, J.M., & Sokolova, Y.Y. (2018). Detection of two Microsporidia pathogens of the European honey bee *Apis mellifera* (Insecta: Apidae) in Western Siberia. *Acta Parasitologica*, 63(4), 728–732. <u>https://doi.org/10.1515/ap-2018-0086</u>
- USSR Ministry of Agriculture (1984). Methodical instructions for express diagnostics of varroatosis and determination of the degree of bee family infestation by varroa mites in apiary conditions. (1984). Approved by the Chief Veterinary Department of the Ministry of Agriculture of the SSSR ot 16.01.1984 № 115-6a.
- Vandervalk, L.P., Nasr, M.E., & Dosdall, L.M. (2014). New miticides for integrated pest management of *Varroa destructor* (Acari: Varroidae) in honey bee colonies on the Canadian prairies. *Journal of Economic Entomology*, 107(6), 2030–2036. https://doi.org/10.1603/EC14048
- Vasileva, M.I., Vorobeva, S.L., & Kokonov, S.I. (2024). Ispol'zovanie fitokompozitsii v pchelovodstve kak ekologicheskii priem stimuliatsii zhiznedeiatel'nosti i immunnogo statusa medonosnykh pchel. Agrarnyi Vestnik Urala, 24(5), 661-669. <u>https://doi.org/10.32417/1997-4868-2024-24-05-661-669</u>
- Vilarem, C., Piou, V., Vogelweith, F., & Vétillard, A. (2021). Varroa destructor from the laboratory to the field: control, biocontrol and IPM perspectives—a review. *Insects*, 12(9), 800. <u>https://doi.org/10.3390/insects12090800</u>
- Wakgari, M., & Yigezu, G. (2021). Honeybee keeping constraints and future prospects. Cogent Food and Agriculture, 7(1). https://doi.org/10.1080/23311932.2021.1872192